# Occurrence, antimicrobial resistance, and potential zoonosis risk of avian pathogenic *Escherichia coli* in Indonesia: A review

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### Abstract

Avian pathogenic Escherichia coli (APEC) causes avian colibacillosis or colibacillosis and is a major endemic disease of poultry worldwide, including in Indonesia. It is characterized by a black proventriculus and can damage other organs, leading to pericarditis, perihepatitis, water sacculitis, mesenteritis, and omphalitis. The APEC strain is one of the six main sub-pathotypes of the extraintestinal pathogenic E. coli (ExPEC) pathotype. The relationship between APEC and infection in humans is questionable. The previous studies have suggested poultry products, including meat and eggs, as a potential source of infection for the transmission of ExPEC disease to humans. Due to the absence of reporting of disease incidents and the lack of literature updates on this disease, it seems as if APEC does not exist in Indonesia. Since bacterial resistance is a growing problem in Indonesia, and globally, the World Health Organization issued a statement regarding the importance of assessing related factors and their control strategies. Antimicrobial resistance, especially multidrug resistance, presents a challenge when treating infectious diseases. In Indonesia, the incidence of resistance to several antimicrobials in cases of avian colibacillosis is high. In addition, avian and human extraintestinal E. coli infections present a potential zoonotic risk. Furthermore, a relationship exists between antibiotic resistance to foodborne bacteria and the occurrence of antibiotic resistance in humans, so the use of antibiotics in the poultry industry must be controlled. Therefore, the One Health strategy should be implemented to prevent the overuse or misuse of antibiotics in the poultry industry. This review aimed to increase awareness of people who are at risk of getting Avian pathogenic Escherichia coli (APEC) from poultry by controlling the spread of APEC by maintaining a clean environment and hygienic personnel in poultry farms.

Keywords: antimicrobial resistance, avian pathogenic *Escherichia coli*, extraintestinal pathogenic *Escherichia coli*, human health, zoonosis.

### Introduction

The poultry industry is the main economic sector in Indonesia [1, 2] and the largest food supplier to the global population. Poultry is preferred over other types of animal food products because it is comparatively cheaper and has a relatively high proportion of animal protein [3–7]. The development of the poultry industry in Indonesia has increased and is driven by the high market demand for poultry commodities. For the majority of the Muslim community, prices that are able to meet purchasing power, fulfillment of animal protein nutrition requirements at all socioeconomic levels, maintenance management, environment, and health are factors that support the success of poultry farming in Indonesia [1, 8, 9]. The relationship among these associated factors must be in balance, and an imbalance of one factor can give rise to a disease. Infectious diseases involve a causative agent and host, as well as environmental factors. Disease is a risk factor that is often faced by all livestock businesses, particularly poultry farming. Therefore, knowledge and information on disease incidence and efforts to prevent, control, and eradicate them are essential [7, 10–13].

Avian pathogenic *Escherichia coli* (APEC), which causes avian colibacillosis in poultry, is considered a major pathogen for the poultry industry. The disease is often named colibacillosis or coli disease by poultry farmers in Indonesia. Serogroups O2 and O78 are the most common in outbreaks of avian colibacillosis, representing 80% of disease cases worldwide [14–18]. Avian pathogenic *E. coli* often occurs in Indonesia [12, 19, 20], and cases of colibacillosis in the field are categorized as secondary diseases that can occur in farms with poor sanitation or secondary infections from respiratory diseases and other immunosuppressive diseases in poultry, such as swollen head syndrome, chronic respiratory disease, and Newcastle disease [3, 21–23].

Although known for more than a century, avian colibacillosis remains one of the main endemic diseases of poultry worldwide, including in Indonesia. Avian

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colibacillosis is a contagious disease in poultry caused by pathogenic *E. coli* bacteria and leads to fairly high economic losses. In Indonesia, this amounts to a loss of 13.10% of the total assets of poultry either directly (weight loss, reduced egg production, and increased total mortality) or indirectly (cleaning, disinfection, and labor compensation) in the event of disease. The proportion of losses incurred to total livestock assets is an indicator of how important it is for the disease to be controlled or overcome [11, 21, 23]. *Escherichia coli* is one of the causes of foodborne zoonoses; it is infectious and produces toxins [19, 24–27].

The incidence of colibacillosis in livestock has led to the use of antibiotics for disease treatment and prevention, and poultry farming is one of the sources of antimicrobial resistance (AMR). The AMR of E. coli is a global public health threat that requires action across all sectors of government and society. Uncontrolled use of antibiotics can lead to antibiotic resistance in layer [28, 29] and broiler [30-32] farms. The high use of antibiotics without a prescription is due to the perception of farmers that their use has no side effects and is a low-cost effort to prevent disease [33]. However, bacterial resistance is a growing problem worldwide, and the World Health Organization issued a statement regarding the importance of assessing the factors associated with this problem and strategies to control the incidence of resistance [34-37]. Antibiotic resistance in one country will now become a problem for all countries [38].

When the incidence of AMR increases, antimicrobial drugs become ineffective, and the infection persists in the body and increases the risk of spreading to the next host. The absence of reporting of disease incidents and the lack of literature updates on this disease make it seem as if APEC does not exist in Indonesia. This review discusses APEC in general, the incidence of APEC in Indonesia, resistance caused by APEC, and the potential risk of APEC as a zoonosis.

# Avian Pathogenic E. coli

Poultry can act as an APEC reservoir which is spread through eggs and meat of chickens and other poultry [39-42]. The digestive tract of a day-old chicken (DOC) broiler is a reservoir of potentially pathogenic E. coli [43]. Avian pathogenic E. coli is a disease in poultry caused by extraintestinal pathogenic E. coli (ExPEC) that causes abnormalities in organs outside the gastrointestinal tract, such as thickened air sacs covered with fibrin fluid (airsacculitis), fibrinous pericarditis, peritonitis, salpingitis, ophthalmia, synovitis, and septicemia. Omphalitis occurs in chicks [17, 44–46]. Avian pathogenic E. coli strains cause primary and secondary diseases in chickens and other poultry breeds [44, 47-49]. In their review article, Filho et al. [50] presented a summary chart of the relationships between pathotypes and sub-pathotypes of E. coli. In this chart, E. coli is divided into two main groups, namely, commensal (nonpathogenic) and

pathogenic *E. coli*. Commensal *E. coli* exists as normal flora in the digestive tract of humans, mammals, and birds and helps the host digest food. It is also involved in competing against other pathogenic bacteria in the digestive tract. Phylogenetically, commensal *E. coli* belongs to *E. coli* Group A and Group B1 and does not possess any virulence factors [50–52].

The APEC strains of *E. coli* produce verotoxin (VT), also known as Shiga toxin (Stx) [19, 53–56]. This cytotoxin is produced by some enteropathogenic *E. coli* (EPEC) strains [57]. Some APEC strains have either VT1 (Stx1) or VT2 (Stx2, airsacculitis) genes or both [54]. Strains with only VT2 (Stx2) are more pathogenic than strains with only VT1 (Stx1) or both [54, 55]. *Escherichia coli* isolates that produce VT1 and VT2 can cause disease in poultry and humans [53].

Pathogenic E. coli is divided into two subgroups according to its predilection, namely, diarrheagenic E. coli (DEC) [54, 58], which is pathogenic in the digestive tract, and ExPEC [26, 59], which causes systemic infections [50]. Diarrheagenic E. coli is found in the intestinal tracts of both humans and animals and is classified into eight sub-pathotypes, including EPEC[60,61], enterotoxigenic E. coli(ETEC)[62-64], enterohemorrhagic Ε. coli (EHEC) [65-67],enteroinvasive E. coli (EIEC) [68-71], diffusely adherent E. coli [72-76], enteroaggregative E. coli (EAEC) [75, 77-81], adherent invasive E. coli [82-85], and Stx-producing EAEC [65, 77, 81, 86]. Diarrheagenic E. coli is transmitted through food or water contaminated with animal or human feces and causes gastroenteritis and diarrhea. Diarrheagenic E. coli transmission by direct human-to-human contact can occur, but this is rare. Among the groups at risk of DEC infection, tourists who have just come to an area can be at greater risk of ETEC infection. Enteropathogenic E. coli and EIEC infections are common in children in developing countries, whereas EAEC infections are common in immunocompromised people. The Stx-producing E. coli (STEC) strain is also known as verotoxigenic E. coli, and the term EHEC is usually used to define STEC strains capable of causing disease in humans, especially bloody diarrhea and hemolytic uremic syndrome [58, 80, 87].

The ExPEC pathotype is divided into six main sub-pathotypes [50, 58], including uropathogenic E. coli (UPEC) [88-92], sepsis/newborn meningitis-associated E. coli (NMEC) [93-96], APEC [45, 97-102], sepsis-associated pathogenic E. coli (SePEC) [103, 104], mammary pathogenic coli [105-108], and endometrial pathogenic Е. E. coli [109, 110]. The ExPEC strain was characterized as E. coli isolate containing two or more virulence factors. Known virulence factors include Type 1 (F1) and P (Pap/Prs) fimbriae for colonization, IbeA for invasion, iron acquisition systems, TraT and Iss for serum survival, K and O antigens for antiphagocytic activity, and temperature-sensitive haemagglutinin [49, 91, 94, 101, 111].

### **Occurrence of APEC in Indonesia**

Avian pathogenic E. coli is an infectious disease in poultry caused by Gram-negative bacteria, including ExPEC strains [26], which lead to diseases outside the intestine, and includes strains that are pathogenic to animals and humans [44, 111]. Avian colibacillosis, characterized by a black proventriculus [112], is a disease that attacks poultry and often occurs in broilers and laying hens. It can infect all ages, from DOCs to the age when they are ready to harvest [40]. However, events in the field indicate that avian colibacillosis more frequently infects chickens at young ages [20, 113]. Colibacillosis is one of the main bacterial diseases that cause high morbidity and mortality in poultry. The most common presentation is a respiratory tract infection, often followed by septicemia [13, 45]. Transmission to healthy chickens occurs through direct contact of infected animals with contaminated drinking water, flies or insects as vectors, and litter or cage mats contaminated with feces from infected chickens. Avian pathogenic E. coli strains can survive for several months in chicken feces [114].

Information on the incidence of disease spread, phylogenetic relationships, host specificity, pathogenicity, and genes associated with the emergence of APEC disease in Indonesia is scarce, and it appears that APEC disease does not exist in Indonesia. We could only final a minimal number of scientific articles reporting APEC incidents in Indonesia [11, 18, 20, 113, 115–117], which indicates a lack of information regarding the incidence of avian colibacillosis in Indonesia, although this disease is known to occur in many Indonesian poultry farms. The high incidence of avian colibacillosis disease in the field has not been widely reported in scientific articles, although in terms of disease losses, it has a fairly high economic impact [22, 100, 111, 118, 119].

# Avian Pathogenic E. coli and AMR

Antimicrobials (antibiotics, disinfectants, and antiseptics) are substances that interfere with the growth and metabolism of microorganisms. The use of these drugs aims to inhibit (bacteriostatic) or kill (bactericidal) microorganisms without damaging the host tissue. Inappropriate antimicrobial use can lead to therapeutic failure and increase the risk of resistance or the occurrence of side effects [120-124]. Since the introduction of penicillin in the mid-20<sup>th</sup> century [125, 126], antibiotics have been used not only in human medicine but also in animal care [127]. Initially, antibiotics were used to treat sick animals, but with the intensification of agriculture, the utilization of antibiotics has expanded to include disease prevention and use as growth promoters [128, 129]. Excessive use of antimicrobials on livestock pollutes the environment and contributes to an increase in resistant microorganisms. This poses a threat not only to human health but also to animal health, animal welfare, and sustainable

livestock production, which has implications for food security and people's livelihoods [37, 130–132]. Antimicrobial abuse renders their use ineffective for both animals and humans because it leads to the development and appearance of AMR in disease-causing microorganisms [133–137]. Antimicrobial resistance prevents antimicrobials from killing or inhibiting bacterial growth, rendering antibiotic therapy for infectious diseases ineffective [121, 122, 138].

The presence of anti bacterial agents, the use of antibiotics, and farmer knowledge are some of the factors that can increase the incidence of antibiotic resistance. Antimicrobial resistance has become a global issue [122, 128, 131, 139-142]. Some literature suggests avian colibacillosis as one of the factors that trigger the high level of antibiotic resistance. A high percentage of AMR to several antibiotics, such as ampicillin, enrofloxacin, tetracycline, erythromycin, trimethoprim-sulfamethoxazole, oxytetracycline, cephalosporin, chloramphenicol, and others, has been reported in cases of avian colibacillosis in Indonesia [20, 143–148]. Multidrug resistance (MDR) refers to the resistance of bacteria to more than 3 classes of antibiotics. The presence of strains with MDR in cases of avian colibacillosis on poultry farms has a high potential for antibiotic therapy failure [31, 141, 149, 150]. Antimicrobial resistance, especially MDR, is a difficult problem to overcome when treating infectious diseases [150].

# Potential Risks of APEC as a Zoonosis

Avian pathogenic E. coli causes disease, particularly in poultry; however, the link between APEC and infection in humans is questionable. Some studies revealed chicken as an ExPEC reservoir for humans [39, 40, 42, 151]. Extraintestinal pathogenic E. coli strains include UPEC in humans and APEC in poultry [41, 45, 100, 111]. The genomic sequences of APEC strain O1:K1:H7 showed similarities with UPEC and NMEC strains in humans. These similarities allow APEC to cause disease in humans, and certain APEC strains have the potential to cause urinary tract infections in humans [17, 93, 152]. The phylogenetic analysis of ExPEC isolates revealed grouping according to genes related to virulence and plasmid replicon type. Extraintestinal pathogenic E. coli isolates were revealed as derived from the pathogenic strain APEC in poultry; NMEC, which causes neonatal meningitis in humans; and UPEC, which causes human urinary tract infections. This grouping implies that it is possible to observe the potential for transmission of certain ExPEC strains between humans and animals as a disease with a zoonotic risk [153, 154]. Poultry products, including meat and eggs, have been reported as a potential source of infection for the transmission of ExPEC disease to humans [41, 151, 155, 156].

Since ExPEC in poultry and humans share several common phylogenetic groups and genes related to their virulence, great attention must be paid to the zoonotic risk of APEC disease [157]. Most cases of ExPEC disease in humans show AMR. Extraintestinal pathogenic *E. coli* strains have the special ability to cause disease in internal organs in humans [25, 58, 158] and other extraintestinal organ infections. Extraintestinal pathogenic *E. coli* strains with AMR are transmitted through contaminated food [44, 155, 156, 159]. A new problem related to ExPEC transmission from food, especially poultry products, is the emergence of diseases with MDR, meaning that APEC creates economic problems as well as animal and human health problems [26, 155].

Avian and human ExPEC infections present a potential zoonotic risk. Extraintestinal pathogenic *E. coli* infections in humans can be systemic (urinary tract infection as a manifestation of UPEC and sepsis that progress to meningitis as a manifestation of NMEC and SePEC), whereas infection in poultry, namely, APEC, can cause airsacculitis, salpingitis, and cellulitis. Both animal and human infections can transfer genes related to virulence and antibiotic resistance, and here, the factors that play a role are poultry products as a source of disease [18, 26, 91, 99, 111, 115, 152].

Foodborne disease has the potential to disrupt animal and human health, including transmitting zoonoses or chemical residues found in foods of animal origin. The AMR of some foodborne bacteria has led to treatment failure of gastrointestinal infections in humans. Foodborne bacteria that show AMR can be transmitted to humans through the food chain or by direct contact. Since there is a relationship between the antibiotic resistance of foodborne bacteria and the occurrence of antibiotic resistance in humans, the use of antibiotics in the livestock industry must be controlled [23, 24, 33, 41, 45, 79, 145, 160–169].

### **One Health Approach for APEC**

The use of antimicrobials is a cost-effective practice to reduce the incidence and mortality rate of avian colibacillosis. However, the overuse and misuse of antibiotics have exerted selective pressure for the emergence of AMR and MDR strains, leading to therapeutic failure and potential economic loss in the poultry industry worldwide. Apart from the spread of antimicrobial-resistant strains, AMR genes can be transferred and disseminated between food-producing animals and human pathogens, and this is a global public health issue [170]. For this reason, our discussion of the presence of APEC isolates may pose two threats: Poor antimicrobial treatment of poultry and the potential risks associated with consumer exposure to poultry products from the food chain. Thus, we also need to elucidate the presence of AMR genes in APEC isolates. This review clearly illustrates that in the context of One Health, genomic analysis can reveal opportunities for timely intervention and prevention of the spread of AMR. Another important One Health strategy is the prevention of infection and disease related to APEC and other poultry pathogens [32, 121, 171, 172] by organizing coordinated scientific and public health

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efforts, ensuring effective surveillance, research, public education, communication, and new policymaking resolutions [173].

#### Conclusion

The poultry sector is the largest supplier of human food. Avian pathogenic E. coli is a poultry disease caused by ExPEC. The main endemic disease of poultry is APEC infection or avian colibacillosis. Poultry acts as an APEC reservoir that is spread to humans through eggs and meat. Many APEC incidents in Indonesia occur in the field, but the lack of reporting and literature updates on this disease makes it seem as if APEC does not exist in Indonesia. The presence of antibacterial agents, the use of antibiotics, and farmer knowledge are some of the factors that can increase the incidence of antibiotic resistance. The problem of AMR has become a global issue. There is a high incidence of AMR to several antibiotics in cases of avian colibacillosis in Indonesia. Overcoming AMR, especially MDR, is challenging when treating infectious diseases. Infection with avian colibacillosis and human ExPEC presents a potential zoonotic risk. Foods of animal origin have the potential to be hazardous to the health of animals and humans, including transmitting zoonoses. There is a relationship between the antibiotic resistance of bacteria present in food of animal origin and the occurrence of antibiotic resistance in humans. Thus, the One Health strategy should be implemented to prevent the overuse or misuse of antibiotics in the poultry industry.

### **Authors' Contributions**

FJW: Conceived the idea and drafted and revised the manuscript. MHE and FMW: Reviewed the manuscript. FJW and MHE: Literature search and edited and reviewed the manuscript. All authors have read and approved the final manuscript.

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### **Competing Interests**

The authors declare that they have no competing interests.

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#### References

1. Ferlito, C. and Respatiadi, H. (2019) Policy Reform on Poultry Industry in Indonesia. Center for Indonesian Policy Studies, Indonesia.

- 2. United States Agency for International Development. (2013) Indonesia's Poultry Value Chain: Costs, Margins, Prices, and Other Issues. United States Agency for International Development, Washington, DC.
- Marangoni, F., Corsello, G., Cricelli, C., Ferrara, N., Ghiselli, A., Lucchin, L. and Poli, A. (2015) Role of poultry meat in a balanced diet aimed at maintaining health and wellbeing: An Italian consensus document. *Food Nutr. Res.*, 59(1): 27606.
- Kralik, G., Kralik, Z., Grčević, M. and Hanžek, D. (2018) Quality of chicken meat. In: Animal Husbandry and Nutrition. IntechOpen, London.
- 5. Aryani, G.A.D. and Jember, I.M. (2019) Analysis of the factors that influence the demand for broiler meat in the province of Bali [Analisis faktor faktor yang mempengaruhi permintaan daging ayam broiler di provinsi Bali]. *E J. EP* Unud., 8(5): 1062–1091.
- Montalvo-Puente, A.P., Torres-Gallo, R., Acevedo-Correa, D., Montero-Castillo, P.M. and Tirado, D.F. (2018) Nutritional comparison of beef, pork and chicken meat from Maracaibo city (Venezuela). *Adv. J. Food Sci. Technol.*, 15(SPL): 218–224.
- Ismail, M., Cahyadi, E.R. and Hardjomidjojo, H. (2019) Risk management of poultry disease in broiler breeders and traders in West Java. J. Manaj. Pengemb. Ind. Kecil Menengah., 14(1): 44–53.
- 8. Departemen Pertanian. (2005) Prospects and Directions of Poultry Agribusiness Development [Prospek Dan Arah Pengembangan Agribisnis Unggas]Pusat Litbang Peternakan, Jakarta.
- Saptana, N., Sayuti, R. and Noekman, K.M. (2016) Poultry Industry: Combining Growth and Equity [Industri Perunggasan: Memadukan Pertumbuhan dan Pemerataan] Forum Penelit. Agro Ekon., 20(1):50.
- Widiasih, D.A. and Budiharta, S. (2012) Epidemiology of zoonoses in Indonesia [Epidemiologi Zoonosis Di Indonesia] Gadjah Mada University Press, Yogyakarta.
- Wibisono, F.M., Wibisono, F.J., Effendi, M.H., Plumeriastuti, H., Hidayatullah, A.R., Hartadi, E.B. and Sofiana, E.D. (2020) A review of salmonellosis on poultry farms: Public health importance. *Syst. Rev. Pharm.*, 11(9): 481–486.
- 12. Wiedosari, E. and Wahyuwardani, S. (2015) A case study on the diseases of broiler chicken in Sukabumi and Bogor districts. *J. Kedokt. Hewan*, 9(1): 9–13.
- Kabir, S.M.L. (2010) Avian colibacillosis and salmonellosis: A closer look at epidemiology, pathogenesis, diagnosis, control and public health concerns. *Int. J. Environ. Res. Public Health*, 7(1): 89–114.
- Knöbl, T., Moreno, A.M., Paixão, R., Gomes, T.A.T., Vieira, M.A.M., da Silva Leite, D., Blanco, J.E. and Ferreira, A.J.P. (2012) Prevalence of avian pathogenic *Escherichia coli* (APEC) clone harboring sfa gene in Brazil. *Scientific World Journal*, 2012(1): 437342.
- 15. Dziva, F., Hauser, H., Connor, T.R., van Diemen, P.M., Prescott, G., Langridge, G.C., Eckert, S., Chaudhuri, R.R., Ewers, C., Mellata, M., Mukhopadhyay, S., Curtiss, R., Dougan, G., Wieler, L.H., Thomson, N.R., Pickard, D.J. and Stevens, M.P. (2013) Sequencing and functional annotation of avian pathogenic *Escherichia coli* serogroup O78 strains reveal the evolution of *E. coli* lineages pathogenic for poultry via distinct mechanisms. *Infect Immun.*, 81(3): 838–849.
- Khoo, L.L., Hasnah, Y., Rosnah, Y., Saiful, N., Maswati, M.A. and Ramlan, M. (2010) The prevalence of avian pathogenic *Escherichia coli* (APEC) in peninsular Malaysia. *Malays. J. Vet. Res.*, 1(1): 27–31.
- 17. Panth, Y. (2019) Colibacillosis in poultry: A review. J. Agric. Nat. Resour., 2(1): 301–311.
- Johnson, T.J., Wannemuehler, Y., Kariyawasam, S., Johnson, J.R., Logue, C.M. and Nolan, L.K. (2012) Prevalence of avian-pathogenic *Escherichia coli* strain O1

genomic islands among extraintestinal and commensal *E. coli* isolates. *J. Bacteriol.*, 194(11): 2846–2853.

- Prihtiyantoro, W., Khusnan, K., Slipranata, M. and Rosyidi, I. (2019) Prevalence of avian pathogenic Escherichia coli (APEC) strains causing colibacillosis in quail [Prevalensi strain avian pathogenic *Escherichia coli* (APEC) penyebab kolibasilosis pada burung puyuh] *J. Sain. Vet.*, 37(1): 69.
- 20. Suryani, A.E., Karimy, M.F., Sofyan, A., Herdian, H. and Wibowo, M.H. (2014) Prevalence in broiler chicken infected by *Escherichia coli* with administration of bio additive, probiotic, and antibiotic. *Widyariset*, 17(2): 233–244.
- Tarmudji, T. (2003) Colibacillosis in Chickens: Etiology, Pathology and Control [Kolibasilosis Pada Ayam: Etiologi, Patologi Dan Pengendaliannya. Wartazoa], 13(2): 65-68.
- 22. Wibowo, M.H. and Wahyuni, A.E.T. (2008) Study of the pathogenicity of Escherichia coli isolates of poultry in 15-day-old broilers [Studi patogenitas *Escherichia coli* isolat unggas pada ayam pedaging umur 15 hari] *J. Vet.*, 9(2): 87–93.
- 23. Wibisono, F.J., Sumiarto, B. and Kusumastuti, T.A. (2018) Economic losses estimation of pathogenic *Escherichia coli* infection in Indonesian poultry farming. *Bul. Peternak.*, 42(4): 341–346.
- 24. De Blackburn, C.W. and McClure, P.J. (2009) Hazards, risk analysis and control woodhead publishing series in food science, technology and nutrition. In: Foodborne Pathogens. 2<sup>nd</sup> ed. Elsevier, Amsterdam, Netherlands. p718–762.
- 25. Wibisono, F.J. (2015) Potensi *Escherichia coli* sebagai foodborne zoonotic disease. *VITEK Bid. Kedokt. Hewan*, 7(7): 55–61.
- 26. Smith, J.L., Fratamico, P.M. and Gunther, N.W. (2007) Extraintestinal pathogenic *Escherichia coli*. *Foodborne Pathog. Dis.*, 4(2): 134–163.
- Mellata, M. (2013) Human and avian extraintestinal pathogenic *Escherichia coli*: Infections, zoonotic risks, and antibiotic resistance trends. *Foodborne Pathog. Dis.*, 10(11): 916–932.
- Wibisono, F.J., Sumiarto, B., Untari, T., Effendi, M.H., Permatasari, D.A. and Witaningrum, A.M. (2020) Short communication: The presence of extended-spectrum beta-lactamase (ESBL) producing *Escherichia coli* on layer chicken farms in Blitar area, Indonesia. *Biodiversitas J. Biol. Divers.*, 21(6): 2667–2671.
- Wibisono, F.J., Sumiarto, B., Untari, T., Effendi, M.H., Permatasari, D.A. and Witaningrum, A.M. (2020) Antimicrobial resistance on *Escherichia coli* from poultry production on Blitar, Indonesia. *Indian J. Forensic. Med. Toxicol.*, 14(4): 4131–4136.
- Effendi, M.H., Wibisono, F.J., Witaningrum, A.M. and Permatasari, D.A. (2021) Identification of Bla TEM and Bla SHV genes of extended spectrum beta lactamase (ESBL) producing *Escherichia coli* from broilers chicken in Blitar, Indonesia. *Syst. Rev. Pharm.*, 12(1): 976–981.
- Wibisono, F.J., Sumiarto, B., Untari, T., Effendi, M.H., Permatasari, D.A. and Witaningrum, A.M. (2020) CTX gene of extended spectrum beta-lactamase (ESBL) producing *Escherichia coli* on broilers in Blitar, Indonesia. *Syst. Rev. Pharm.*, 11(7): 396–403.
- 32. Effendi, M.H., Tyasningsih, W., Yurianti, Y.A., Rahmahani, J., Harijani, N. and Plumeriastuti, H. (2021) Presence of multidrug resistance (MDR) and extended-spectrum beta-lactamase (ESBL) of *Escherichia coli* isolated from cloacal swabs of broilers in several wet markets in Surabaya, Indonesia. *Biodiversitas*, 22(1): 304–310.
- 33. Niasono, A.B., Latif, H. and Purnawarman, T. (2019) Antibiotic resistance to *Escherichia coli* bacteria isolated from broiler farms in Subang district, West Java [Resistensi antibiotik terhadap bakteri *Escherichia coli* yang diisolasi dari peternakan ayam pedaging di kabupaten subang, Jawa Barat] J. Vet., 20(36): 187–195.
- 34. Utami, E.R. (2011) Antibiotics, resistance, and rationality

of therapy [Antibiotika, resistensi, dan rasionalitas terapi] *El Haya*h., 1(4): 3.

- Thanner, S., Drissner, D. and Walsh, F. (2016) Antimicrobial resistance in agriculture. *Am. Soc. Microbiol.*, 7(2): 1–7.
- Iwu, C.D., Korsten, L. and Okoh, A.I. (2020) The incidence of antibiotic resistance within and beyond the agricultural ecosystem: A concern for public health. *Microbiologyopen*, 9(9): 1–28.
- 37. Wall, B.A., Mateus, A., Marshall, L., Pfeiffer, D., Lubroth, J., Ormel, H.J., Otto, P. and Patriarchi, A. (2016) The emergence of antimicrobial resistance in bacteria. In: Drivers, Dynamics and Epidemiology of Antimicrobial Resistance in Animal Production. Veterinary Epidemiology, Economics and Public Health Group, Department of Production and Population Health, The Royal Veterinary College, North Mymms, London, UK. p1–58.
- Aarestrup, F.M. (2015) The livestock reservoir for antimicrobial resistance: A personal view on changing patterns of risks, effects of interventions and the way forward. *Philos. Trans. R. Soc. Lond. B Biol. Sci.*, 370(1670): 20140085.
- Radwan, I.A.E., Sayed, H., Salam, H. and Abdalwanis, S.A.A. (2014) Frequency of some virulence associated genes among multidrug-resistant *Escherichia coli* isolated from septicemic broiler chicken. *Int. J. Adv. Res.*, 2(12): 867–874.
- Mora, A., Viso, S., López, C., Alonso, M.P., García-Garrote, F., Dabhi, G., Mamani, R., Herrera, A., Marzoa, J., Blanco, M., Blanco, J.E., Moulin-Schouleur, M., Schouler, C. and Blanco, J. (2013) Poultry as reservoir for extraintestinal pathogenic *Escherichia coli* O45: K1: H7-B2-ST95 in humans. *Vet. Microbiol.*, 167(3–4): 506–512.
- Kemmett, K., Humphrey, T., Rushton, S., Close, A., Wigley, P. and Williams, N.J. (2013) A longitudinal study simultaneously exploring the carriage of APEC virulence associated genes and the molecular epidemiology of faecal and systemic *E. coli* in commercial broiler chickens. *PLoS One.*, 8(6): e67749.
- 42. Mitchell, N.M., Johnson, J.R., Johnston, B., Curtiss, R. and Mellata, M. (2015) Zoonotic potential of *Escherichia coli* isolates from retail chicken meat products and eggs. *Appl. Environ. Microbiol.*, 81(3): 1177–1187.
- Kemmett, K., Williams, N.J., Chaloner, G., Humphrey, S., Wigley, P. and Humphrey, T. (2014) The contribution of systemic *Escherichia coli* infection to the early mortalities of commercial broiler chickens. *Avian Pathol.*, 43(1): 37–42.
- Dho-moulin, M. and Fairbrother, J.M. (1999) Avian pathogenic *Escherichia coli* (APEC). *Vet. Res.*, 30(2–3): 299–316.
- 45. Stromberg, Z.R., Johnson, J.R., Fairbrother, J.M., Kilbourne, J., Van Goor, A., Curtiss, R. and Mellata, M. (2017) Evaluation of *Escherichia coli* isolates from healthy chickens to determine their potential risk to poultry and human health. *PLoS One*, 12(7): 1–18.
- Kabir, S.M.L., Sikder, M.H., Alam, J., Neogi, S.B. and Yamasaki, S. (2017) Colibacillosis and its impact on egg production. In: Egg Innovations and Strategies for Improvements. Elsevier Inc., Amsterdam, Netherlands. p523–535.
- 47. Antão, E.M., Glodde, S., Li, G., Sharifi, R., Homeier, T., Laturnus, C., Diehl, I., Bethe, A., Philipp, H.C., Preisinger, R., Wieler, L.H. and Ewers, C. (2008) The chicken as a natural model for extraintestinal infections caused by avian pathogenic *Escherichia coli* (APEC). *Microb. Pathog.*, 45(5–6): 361–369.
- Ewers, C., Janssen, T. and Wieler, L. (2003) Avian pathogenic *Escherichia coli* (APEC). *Berl. Munch. Tierarztl. Wochenschr.*, 30(2–3): 299–316.
- Dziva, F. and Stevens, M.P. (2008) Colibacillosis in poultry: Unravelling the molecular basis of virulence of avian pathogenic *Escherichia coli* in their natural hosts. *Avian Pathol.*, 37(4):355–366.
- International Journal of One Health, EISSN: 2455-8931

- Filho, H.C.K., Brito, K.C.T., Cavalli, L.S. and Brito, B.G. (2015) Avian pathogenic *Escherichia coli* (APEC) an update on the control. In: The Battle against Microbial Pathogens: Basic Science, Technological Advances and Educational Programs. Formatex Research Center. p598–618.
- Duriez, P., Clermont, O., Bonacorsi, S., Bingen, E., Chaventré, A., Elion, J., Picard, B. and Denamur, E. (2001) Commensal *Escherichia coli* isolates are phylogenetically distributed among geographically distinct human populations. *Microbiology*, 147(6): 1671–1676.
- Bailey, J.K., Pinyon, J.L., Anantham, S. and Hall, R.M. (2010) Distribution of human commensal *Escherichia coli* phylogenetic groups. *J. Clin. Microbiol.*, 48(9): 3455–3456.
- Saikia, M.K. and Saikia, D. (2011) PCR detection of "STX1" AND "STX2" Toxigenic genes in multiple antibiotic resistant *Escherichia coli* population and phenotypic detection of ESBL producing *Escherichia coli* isolates from local variety of poultry. *Int. J. Appl. Biol. Pharm. Technol.*, 2(3): 593–602.
- 54. Nataro, J.P. and Kaper, J.B. (1998) Diarrheagenic *Escherichia coli. Clin. Microbiol. Rev.*, 11(1): 142–201.
- 55. Ludwig, K., Sarkim, V., Bitzan, M., Karmali, M.A., Bobrowski, C., Ruder, H., Laufs, R., Sobottka, I., Petric, M., Karch, H. and Müller-Wiefel, D.E. (2002) Shiga toxin-producing *Escherichia coli* infection and antibodies against Stx2 and Stx1 in household contacts of children with enteropathic hemolytic-uremic syndrome. *J. Clin. Microbiol.*, 40(5): 1773–1782.
- Kwon, H.J., Seong, W.J. and Kim, J.H. (2013) Molecular prophage typing of avian pathogenic *Escherichia coli*. *Vet. Microbiol.*, 162(2–4): 785–792.
- 57. Salehi, T.Z., Safarchi, A., Peighambari, S.M., Mahzounieh, M. and Khorasgani, M.R. (2007) Detection of stx1, stx2, eae, esp B and hly genes in avian pathogenic *Escherichia coli* by multiplex polymerase chain reaction. J. *Vet. Res.*, 62(2): 37–42.
- O'Reilly, C.E., Iwamoto, M. and Griffin, P.M. (2017) *Escherichia coli*, Diarrheagenic. Centers for Disease Control and Prevention, Atlanta, Georgia.
- Dale, A.P. and Woodford, N. (2015) Extra-intestinal pathogenic *Escherichia coli* (ExPEC): Disease, carriage and clones. J. Infect., 71(6): 615–626.
- Ochoa, T.J. and Contreras, C.A. (2011) Enteropathogenic Escherichia coli infection in children. Curr. Opin. Infect. Dis., 24(5): 478–483.
- 61. Pearson, J.S. and Frankel, G. (2016) Immunity to enteropathogenic *Escherichia coli*. In: Encyclopedia of Immunobiology. Elsevier, Amsterdam, Netherlands. p43–51.
- Evans, D. and Evans, D. (1996) Medical microbiology. In: Baron S, editor. *Escherichia Coli* in Diarrheal Disease: Medical Microbiology. 4<sup>th</sup> ed., Ch. 25. Texas: Texas Medical Branch at Galveston, Galveston, Texas.
- Crofts, A.A., Giovanetti, S.M., Rubin, E.J., Poly, F.M., Gutiérrez, R.L., Talaat, K.R., Porter, C.K., Riddle, M.S., DeNearing, B., Brubaker, J., Maciel, M., Alcala, A.N., Chakraborty, S., Prouty, M.G., Savarino, S.J., Davies, B.W. and Trent, M.S. (2018) Enterotoxigenic *E. coli* virulence gene regulation in human infections. *Proc. Natl. Acad. Sci.* U. S. A., 115(38): E8968–E8976.
- Fleckenstein, J.M. (2013) Enterotoxigenic *Escherichia coli*. In: *Escherichia coli*: Pathotypes and Principles of Pathogenesis. 2<sup>nd</sup> ed. Elsevier, Amsterdam, Netherlands. p183–213.
- Page, A.V. and Liles, W.C. (2013) Enterohemorrhagic Escherichia coli infections and the hemolytic-uremic syndrome. Med. Clin. North Am., 97(4): 681–695.
- 66. LeStrange, K., Markland, S.M., Hoover, D.G., Sharma, M. and Kniel, K.E. (2017) An evaluation of the virulence and adherence properties of avian pathogenic *Escherichia coli*. *One Health*, 4(1): 22–26.
- 67. Fatima, R. and Aziz, M. (2021) Enterohemorrhagic

*Escherichia Coli*. Treasure Island, FL: StatPearls Publishing. p1–5.

- Belotserkovsky, I. and Sansonetti, P.J. (2018) Shigella and enteroinvasive *Escherichia coli. Curr. Top. Microbiol. Immunol.*, 416(1): 1–26.
- 69. Tonu, N.S., Sufian, M.A., Sarker, S., Kamal, M.M., Rahman, M.H. and Hossain, M.M. (2011) Pathological study on colibacillosis in chickens and detection of *Escherichia coli* by Pcr. *Bangladesh J. Vet. Med.*, 9(1): 17–25.
- 70. Michelacci, V., Tozzoli, R., Arancia, S., D'Angelo, A., Boni, A., Knijn, A., Prosseda, G., Greig, D.R., Jenkins, C., Camou, T., Sirok, A., Navarro, A., Schelotto, F., Varela, G. and Morabito, S. (2020) Tracing back the evolutionary route of enteroinvasive *Escherichia coli* (EIEC) and *Shigella* through the example of the highly pathogenic O96:H19 EIEC clone. *Front. Cell. Infect. Microbiol.*, 10(1): 260.
- Farajzadeh-Sheikh, A., Savari, M., Ahmadi, K., Hosseini, N.H., Shahin, M. and Afzali, M. (2020) Distribution of genes encoding virulence factors and the genetic diversity of Enteroinvasive *Escherichia coli* (EIEC) isolates from patients with diarrhea in Ahvaz, Iran. *Infect. Drug Resist.*, 13(1): 119–127.
- Scaletsky, I.C.A., Fabbricotti, S.H., Carvalho, R.L.B., Nunes, C.R., Maranhão, H.S., Morais, M.B. and Fagundes-Neto, U. (2002) Diffusely adherent *Escherichia coli* as a cause of acute diarrhea in young children in Northeast Brazil: A case-control study. *J. Clin. Microbiol.*, 40(2): 645–648.
- Javadi, K., Mohebi, S., Motamedifar, M. and Hadi, N. (2020) Characterization and antibiotic resistance pattern of diffusely adherent *Escherichia coli* (DAEC), isolated from paediatric diarrhoea in Shiraz, Southern Iran. *New Microbes New Infect.*, 38(1): 100780.
- Abbasi, P., Kargar, M., Doosti, A., Mardaneh, J., Ghorbani-Dalini, S. and Dehyadegari, M.A. (2017) Molecular detection of diffusely adherent *Escherichia coli* strains associated with diarrhea in Shiraz, Iran. *Arch Pediatr. Infect. Dis.*, 5(2): e37629.
- 75. Meza-Segura, M., Zaidi, M.B., Vera-Ponce de León, A., Moran-Garcia, N., Martinez-Romero, E., Nataro, J.P. and Estrada-Garcia, T. (2020) New insights into DAEC and EAEC pathogenesis and phylogeny. *Front Cell. Infect. Microbiol.*, 10(1): 572951.
- Le Bouguénec, C. and Servin, A.L. (2006) Diffusely adherent *Escherichia coli* strains expressing Afa/Dr adhesins (Afa/Dr DAEC): Hitherto unrecognized pathogens. *FEMS Microbiol. Lett.*, 256(2): 185–194.
- Jandhyala, D.M., Vanguri, V., Boll, E.J., Lai, Y.S., McCormick, B.A. and Leong, J.M. (2013) Shiga toxin-producing *Escherichia coli* O104: H4. An emerging pathogen with enhanced virulence. *Infect. Dis. Clin. North Am.*, 27(3): 631–649.
- Kaur, P., Chakraborti, A. and Asea, A. (2010) Enteroaggregative *Escherichia coli*: An emerging enteric food borne pathogen. *Interdiscip. Perspect. Infect. Dis.*, 2010(1): 254159.
- 79. Ellis, S.J., Crossman, L.C., McGrath, C.J., Chattaway, M.A., Hölken, J.M., Brett, B., Bundy, L., Kay, G.L., Wain, J. and Schüller, S. (2020) Identification and characterisation of enteroaggregative *Escherichia coli* subtypes associated with human disease. *Sci. Rep.*, 10(1): 1–12.
- Brüssow, H. (2014) *Escherichia coli*: Enteroaggregative *E. coli*. In: Encyclopedia of Food Microbiology. 2<sup>nd</sup> ed., Vol. 1. Elsevier, Amsterdam, Netherlands. p706–712.
- Kimata, K., Lee, K., Watahiki, M., Isobe, J., Ohnishi, M. and Iyoda, S. (2020) Global distribution of epidemic-related Shiga toxin 2 encoding phages among enteroaggregative *Escherichia coli. Sci. Rep.*, 10(1): 1–8.
- Chervy, M., Barnich, N. and Denizot, J. (2020) Adherentinvasive *E. coli*: Update on the lifestyle of a troublemaker in Crohn's disease. *Int. J. Mol. Sci.*, 21(10): 1–34.
- 83. Petersen, A.M., Nielsen, E.M., Litrup, E., Brynskov, J.,

Mirsepasi, H. and Krogfelt, K. (2009) A phylogenetic group of *Escherichia coli* associated with active left-sided inflammatory bowel disease. *BMC Microbiol.*, 9(1): 171.

- Lee, J.G., Han, D.S., Jo, S.V., Reum Lee, A., Park, C.H., Eun, C.S. and Lee, Y. (2019) Characteristics and pathogenic role of adherent-invasive *Escherichia coli* in inflammatory bowel disease: Potential impact on clinical outcomes. *PLoS One*, 14(4): 1–14.
- Camprubi-Font, C. and Martinez-Medina, M. (2020) Why the discovery of adherent-invasive *Escherichia coli* molecular markers is so challenging? *World J. Biol. Chem.*, 11(1): 1–13.
- Effendi, M.H., Harijani, N., Yanestria, S.M. and Hastutiek, P. (2018) Identification of Shiga toxin-producing *Escherichia coli* in raw milk samples from dairy cows in Surabaya, Indonesia. *Philipp. J. Vet. Med.*, 55(SI): 109–114.
- 87. Moriel, D.G., Rosini, R., Seib, K.L., Serino, L., Pizza, M. and Rappuoli, R. (2012) *Escherichia coli*: Great diversity around a common core. *mBio*, 3(3): 6–8.
- Antão, E.M., Wieler, L.H. and Ewers, C. (2009) Adhesive threads of extraintestinal pathogenic *Escherichia coli. Gut. Pathog.*, 1(1): 22.
- Shah, C., Baral, R., Bartaula, B. and Shrestha, L.B. (2019) Virulence factors of uropathogenic *Escherichia coli* (UPEC) and correlation with antimicrobial resistance. *BMC Microbiol.*, 19(1): 1–6.
- Terlizzi, M.E., Gribaudo, G. and Maffei, M.E. (2017) UroPathogenic *Escherichia coli* (UPEC) infections: Virulence factors, bladder responses, antibiotic, and non-antibiotic antimicrobial strategies. *Front Microbiol.*, 8(1): 1566.
- 91. Rezatofighi, S.E., Mirzarazi, M. and Salehi, M. (2021) Virulence genes and phylogenetic groups of uropathogenic *Escherichia coli* isolates from patients with urinary tract infection and uninfected control subjects: A case-control study. *BMC Infect. Dis.*, 21(1): 1–11.
- 92. Kot, B., Grużewska, A., Szweda, P., Wicha, J. and Parulska, U. (2020) Antibiotic resistance of uropathogens isolated from patients hospitalized in district hospital in central Poland in 2020. *Antibiotics (Basel)*, 10(4): 447.
- 93. Kim, K.S. (2016) Human meningitis-associated *Escherichia coli. EcoSal Plus*, 7(1): 269–286.
- 94. Breland, E.J., Eberly, A.R. and Hadjifrangiskou, M. (2017) An overview of two-component signal transduction systems implicated in extra-intestinal pathogenic *E. coli* infections. *Front. Cell. Infect. Microbiol.*, 7(1): 162.
- 95. Vissing, N.H., Mønster, M.B., Nordly, S., Dayani, G.K., Heedegaard, S.S., Knudsen, J.D. and Nygaard, U. (2021) Relapse of neonatal *Escherichia coli* meningitis: Did we miss something at first? *Children (Basel)*, 8(2): 126.
- Wijetunge, D.S.S., Gongati, S., Debroy, C., Kim, K.S., Couraud, P.O., Romero, I.A., Weksler, B. and Kariyawasam, S. (2015) The pathotype of neonatal meningitis causing *Escherichia coli* (NMEC) applied microbiology. *BMC Microbiol.*, 15(1): 1–15.
- 97. Alber, A., Morris, K.M., Bryson, K.J., Sutton, K.M., Monson, M.S., Chintoan-Uta, C., Borowska, D., Lamont, S.J., Schouler, C., Kaiser, P., Stevens, M.P. and Vervelde, L. (2020) Avian pathogenic *Escherichia coli* (APEC) strain-dependent immunomodulation of respiratory granulocytes and mononuclear phagocytes in CSF1Rreporter transgenic chickens. *Front. Immunol.*, 10(1): 3055.
- Ronco, T., Stegger, M., Olsen, R.H., Sekse, C., Nordstoga, A.B., Pohjanvirta, T., Lilje, B., Lyhs, U., Andersen, P.S. and Pedersen, K. (2017) Spread of avian pathogenic *Escherichia coli* ST117 O78:H4 in Nordic broiler production. *BMC Genomics*, 18(1): 13.
- 99. Silveira, F., Maluta, R.P., Tiba, M.R., de Paiva, J.B., Guastalli, E.A.L. and da Silveira, W.D. (2016) Comparison between avian pathogenic (APEC) and avian faecal (AFEC) *Escherichia coli* isolated from different regions in Brazil. *Vet. J.*, 217(1): 65–67.

- Collingwood, C., Kemmett, K., Williams, N. and Wigley, P. (2014) Is the concept of avian pathogenic *Escherichia coli* as a single pathotype fundamentally flawed? *Front. Vet. Sci.*, 1(1): 5.
- 101. Cummins, M.L., Reid, C.J., Chowdhury, P.R., Bushell, R.N., Esbert, N., Tivendale, K.A., Noormohammadi, A.H., Islam, S., Marenda, M.S., Browning, G.F., Markham, P.F. and Djordjevic, S.P. (2019) Whole genome sequence analysis of Australian avian pathogenic *Escherichia coli* that carry the class 1 integrase gene. *Microb. Genom.*, 5(2): e000250.
- 102. Yi, Z., Wang, D., Xin, S., Zhou, D., Li, T., Tian, M., Qi, J., Ding, C., Wang, S. and Yu, S. (2019) The CpxR regulates Type VI secretion system 2 expression and facilitates the interbacterial competition activity and virulence of avian pathogenic *Escherichia coli*. *Vet. Res.*, 50(1): 40.
- 103. Conceição, R.A., Ludovico, M.S., Andrade, C.G.T. and Yano, T. (2012) Human sepsis-associated *Escherichia coli* (SEPEC) is able to adhere to and invade kidney epithelial cells in culture. *Brazilian J. Med. Biol. Res.*, 45(5): 417–424.
- 104. Kumar, A., Leite, A.F.V., Maekawa, L.S., Kaur, R., Filo, S.J.B., Persaud, P., Shaikh, J.D., Kichloo, A. and Shiwalkar, N. (2020) Management of *E. coli* sepsis. In: *E. coli* Infections importance of Early Diagnosis and Efficient Treatment. IntechOpen, London. p1–29.
- 105. Blum, S.E., Heller, D.E., Jacoby, S., Krifuks, O., Merin, U., Silanikove, N., Lavon, Y. and Leitner, G. (2020) Physiological response of mammary glands to *Escherichia coli* infection: A conflict between glucose need for milk production and immune response. *Sci. Rep.*, 10(1): 1–14.
- 106. Blum, S.E., Goldstone, R.J., Connolly, J.P.R., Répérant-Ferter, M., Germon, P., Inglis, N.F., Krifucks, O., Mathur, S., Manson, E., Mclean, K., Rainard, P., Roe, A.J., Leitner, G. and Smith, D.G.E. (2018) Postgenomics characterization of an essential genetic determinant of mammary pathogenic *Escherichia coli. mBio*, 9(2): e00423–18.
- 107. Blum, S.E., Heller, E.D., Sela, S., Elad, D., Edery, N. and Leitner, G. (2015) Genomic and phenomic study of mammary pathogenic *Escherichia coli*. *PLoS One*, 10(9): 1–24.
- Shpigel, N.Y., Elazar, S. and Rosenshine, I. (2008) Mammary pathogenic *Escherichia coli*. *Curr. Opin. Microbiol.*, 11(1): 60–65.
- 109. Sheldon, I.M., Rycroft, A.N., Dogan, B., Craven, M., Bromfield, J.J., Chandler, A., Roberts, M.H., Price, S.B., Gilbert, R.O. and Simpson, K.W. (2010) Specific strains of *Escherichia coli* are pathogenic for the endometrium of cattle and cause pelvic inflammatory disease in cattle and mice. *PLoS One*, 5(2): e9192.
- 110. Lopes, C.E., De Carli, S., Weber, M.N., Fonseca, A.C.V., Tagliari, N.J., Foresti, L., Cibulski, S.P., Mayer, F.Q., Canal, C.W. and Siqueira, F.M. (2020) Insights on the genetic features of endometrial pathogenic *Escherichia coli* strains from pyometra in companion animals: Improving the knowledge about pathogenesis. *Infect. Genet. Evol.*, 85(1): 104453.
- 111. Maciel, J.F., Matter, L.B., Trindade, M.M., Camillo, G., Lovato, M., de Ávila Botton, S. and de Vargas, A.C. (2017) Virulence factors and antimicrobial susceptibility profile of extraintestinal *Escherichia coli* isolated from an avian colisepticemia outbreak. *Microb. Pathog.*, 103(1): 119–122.
- 112. Cunha, M.P.V., Saidenberg, A.B., Moreno, A.M., Ferreira, A.J.P., Vieira, M.A.M., Gomes, T.A.T. and Knöbl, T. (2017) Pandemic extra-intestinal pathogenic *Escherichia coli* (ExPEC) clonal group O6-B2-ST73 as a cause of avian colibacillosis in Brazil. *PLoS One*, 12(6): 1–11.
- 113. Wang, X., Cao, C., Huan, H., Zhang, L., Mu, X., Gao, Q., Dong, X., Gao, S. and Liu, X. (2015) Isolation, identification, and pathogenicity of O142 avian pathogenic *Escherichia coli* causing black proventriculus and septicemia in broiler breeders. *Infect. Genet. Evol.*, 32(1): 23–29.

- Santoso, S.W.H., Ardana, I.B.K. and Gelgel, K.T.P. (2020) Prevalensi colibacillosis pada broiler yang diberi pakan tanpa antibiotic growth promoters. *Indones. Med. Vet.*, 9(2): 197–205.
- 115. Teplitski, M.E. (2009) *E. coli* and *Salmonella* on Animal Farms: Sources, Survival and Management. The Soil and Water Science Department, Florida.
- Niyatri. (2015) Kejadian Penyakit Colibacillosis Pada Ayam Layer Di Pt. Janu Putra Sejahtera Yogyakarta, Yogyakarta.
- 117. Utomo, B.N. (1996) Kkolibasilosis pada ayam buras di Kalimantan selatan. *Bul. FKH UGM.*, 15(1): 15–28.
- Purnomo, S. and Juarini, E. (1996) Penyebaran Escherichia coli Serotipe O1K1, 02K1 dan 078K80 pada Ayam di Indonesia. J. Ilmu Ternak. Vet. 1(3): 194–199.
- Landman, W.J.M. and van Eck, J.H.H. (2015) The incidence and economic impact of the *Escherichia coli* peritonitis syndrome in Dutch poultry farming. *Avian Pathol.*, 44(5): 370–378.
- 120. Mbanga, J. and Nyararai, Y.O. (2015) Virulence gene profiles of avian pathogenic *Escherichia coli* isolated from chickens with colibacillosis in Bulawayo, Zimbabwe. *Onderstepoort J. Vet. Res.*, 82(1): E1–E8.
- 121. Permatasari, D.A., Witaningrum, A.M., Wibisono, F.J. and Effendi, M.H. (2020) Detection and prevalence of multidrug-resistant *Klebsiella pneumoniae* strains isolated from poultry farms in Blitar, Indonesia. *Biodiversitas*, 21(10): 4642–4647.
- 122. Putra, A.R., Effendi, M.H., Koesdarto, S., Suwarno, S., Tyasningsih, W. and Estoepangestie, A.T. (2020) Detection of the extended spectrum β-lactamase produced by *Escherichia coli* from dairy cows by using the vitek-2 method in Tulungagung Regency, Indonesia. *Iraqi J. Vet. Sci.*, 34(1): 203–207.
- 123. World Health Organization. (2014) Antimicrobial Resistance: Global Report on Surveillance. WHO Library Cataloguing-in-publication Data Antimicrobial France. World Health Organization, Geneva.
- 124. Ansharieta, R., Effendi, M.H. and Plumeriastuti, H. (2020) Detection of multidrug-resistant (MDR) *Escherichia coli* isolated from raw milk in East Java Province, Indonesia. *Indian J. Forensic. Med. Toxicol.*, 14(4): 3403–3407.
- 125. Herwanadan, E. (2007) Resistance of enteric bacteria: Global aspects of antimicrobials [Resistensi dari bakteri enterik: Aspek global terhadap antimikroba] Univ. Med., 26(1): 46–56.
- 126. Lobanovska, M. and Pilla, G. (2017) Penicillin's discovery and antibiotic resistance: Lessons for the future. *Yale J. Biol. Med.*, 90(1): 135–145.
- 127. Gaynes, R. (2017) The discovery of penicillin new insights after more than 75 years of clinical use. *Emerg. Infect. Dis.*, 23(5): 849–853.
- 128. Gustafson, R.H. and Bowen, R.E. (1997) Antibiotic use in animal agriculture. J. Appl. Microbiol., 83(5): 531–541.
- Van, T.T.H., Yidana, Z., Smooker, P.M. and Coloe, P.J. (2020) Antibiotic use in food animals worldwide, with a focus on Africa: Pluses and minuses. J. Glob. Antimicrob. Resist., 20(1): 170–177.
- 130. Brown, K., Uwiera, R.R.E., Kalmokoff, M.L., Brooks, S.P.J. and Inglis, G.D. (2017) Antimicrobial growth promoter use in livestock: A requirement to understand their modes of action to develop effective alternatives. *Int. J. Antimicrob. Agents*, 49(1): 12–24.
- Landers, T.F., Cohen, B., Wittum, T.E. and Larson, E.L. (2012) A review of antibiotic use in food animals: Perspective, policy, and potential. *Public Health Rep.*, 127(1): 4–22.
- 132. Di Martino, G., Crovato, S., Pinto, A., Dorotea, T., Mascarello, G., Brunetta, R., Agnoletti, F. and Bonfanti, L. (2019) Farmers' attitudes towards antimicrobial use and awareness of antimicrobial resistance: A comparative study among Turkey and rabbit farmers. *Ital. J. Anim. Sci.*, 18(1): 194–201.

- 133. Negara, K.S. (2014) Analysis of policy implementation on rational use of antibiotics to prevent antibiotic resistance at Sanglah General Hospital, Denpasar: Case study of methicillin-resistant Staphylococcus aureus infection analysis [Analisis implementasi kebijakan penggunaan antibiotika rasional untuk mencegah resistensi antibiotika di RSUP sanglah denpasar: Studi kasus infeksi methicillin resistant *Staphylococcus aureus* analysis. The implementation policy of rational use of antibiot] *JARS*I, 1(1): 42–50.
- 134. Food and Agriculture Organization. (2018) Animal Health: Antimicrobial Resistance. FAO Departments and Offices. Food and Agriculture Organization of the United Nations, Rome, Italy.
- Manyi-Loh, C., Mamphweli, S., Meyer, E. and Okoh, A. (2018) Antibiotic use in agriculture and its consequential resistance in environmental sources: Potential public health implications. *Molecules*, 23(4): 795.
- Uchil, R.R., Kohli, G.S., Katekhaye, V.M. and Swami, O.C. (2014) Strategies to combat antimicrobial resistance. J. Clin. Diagn. Res., 8(7): 8–11.
- 137. Mouhieddine, T.H., Olleik, Z., Itani, M.M., Kawtharani, S., Nassar, H., Hassoun, R., Houmani, Z., El Zein, Z., Fakih, R., Mortada, I.K., Mohsen, Y., Kanafani, Z. and Tamim, H. (2015) Assessing the Lebanese population for their knowledge, attitudes and practices of antibiotic usage. J. Infect. Public Health, 8(1): 20–31.
- 138. Saliu, E.M., Vahjen, W. and Zentek, J. (2017) Types and prevalence of extended-spectrum beta-lactamase producing *Enterobacteriaceae* in poultry. *Anim. Health Res. Rev.*, 18(1): 46–57.
- Permenkes. (2011) Pedoman Umum Penggunaan Antibiotik. Peraturan Menteri Kesehatan No 2406/MENKES/PER/ XII/2011, Indonesia.
- 140. Estiningsih, D., Puspitasari, I. and Nuryastuti, T. (2016) Identification of multidrug-resistant organisms infection in neonatal intensive care unit (UNIT). *J. Manaj. Pelayanan Farm.*, 6(1): 243–248.
- 141. Rukmini, R., Siahaan, S. and Sari, I.D. (2019) Analysis of the implementation of the antimicrobial control program policy [Analisis implementasi kebijakan program pengendalian antimikroba (PPRA)] Bul. Penelitian Sist. Kesehatan, 22(2): 106–116.
- 142. Wibisono, F.J., Sumiarto, B., Untari, T., Effendi, M.H., Permatasari, D.A. and Witaningrum, A.M. (2020) Prevalence and risk factors analysis of multidrug resistance of *Escherichia coli* bacteria in commercial chicken, Blitar district. *J. Ilmu Peternak Vet. Trop.*, 10(1): 15.
- 143. Purbo, W.A. (2014) Identifikasi Faktor Risiko Terjadinya Resistensi *Enterobacteriaceae* Pada Daging Ayam Broiler dan Ayam Lokal di Kabupaten Bogor. Institute Pertanian Bogor, Bogor.
- 144. Krisnaningsih, M.M.F., Asmara, W. and Wibowo, M.H. (2005) Sensitivity test of pathogenic *Escherichia coli* isolates in chicken to several types of antibiotics [ji sensitivitas isolat *Escherichia coli* patogen pada ayam terhadap beberapa jenis antibiotik] *J. Sain Vet.*, 23(1): 13–18.
- 145. Nugroho, W.S. and Wibowo, M.H. (2005) Sensitivity test of Escherichia coli bacteria isolated from chicken that reacted positively on Congo Red media to ampicillin, streptomycin, and enrofloxacin preparations [Uji sensitivitas bakteri *Escherichia col*i isolat asal ayam yang bereaksi positif pada media Congo Red terhadap preparat ampisilin, streptomicin, dan enrofloksasin] *J. Sain Vet.*, 23(1): 19–23.
- 146. Noor, S.M. and Poeloengan, M. (2005) Use of antibiotics in livestock and their impact on human health [Pemakaian antibiotika pada ternak dan dampaknya pada Kesehatan Manusia] Lokakarya Nas Keamanan Pangan Prod Peternak. p56–64.
- 147. Suharsa, I.W.A., Suarjana, G.K. and Gelgel, K.T.P. (2015) Sensitivity pattern of Escherichia coli isolated from broiler faeces with diarrhea to sulfamethoxazole, ampicillin and oxytetracycline [Pola kepekaan *Escherichia coli* yang

diisolasi dari feses broiler penderita diare terhadap sulfametoksazol, ampisilin dan oksitetrasiklin] *Bul. Vet. Udayana*, 7(2): 101–106.

- 148. Susanto, E. (2014) Escherichia coli which is resistant to antibiotics isolated from broiler chickens and local chickens in Bogor Regency Escherichia coli Yang Resisten Terhadap Antibiotik Yang Diisolasi Dari Ayam Broiler dan Ayam Lokal di Kabupaten Bogor] Institut Pertanian Bogor, Bogor.
- 149. Puspitasari, Y., Handijatno, D. and Ratnasari, R. (2015) *Escherichia coli* resistance test as the cause of colibacillosis in chickens to several types of antibiotics [Uji resistensi *Escherichia coli* sebagai penyebab kolibasilosis pada ayam terhadap beberapa jenis antibiotik] *J. Agro Vet.*, 4(1): 69–76.
- Matin, M.A., Islam, M.A. and Khatun, M.M. (2017) Prevalence of colibacillosis in chickens in greater Mymensingh district of Bangladesh. *Vet. World*, 10(1): 29–33.
- 151. Kurniawati, A.F., Satyabakti, P. and Arbianti, N. (2015) Perbedaan risiko multidrug resistance organisms (MDROS) menurut faktor risiko dan kepatuhan hand hygiene. *J. Berk. Epidemiol.*, 3(3): 277–289.
- Bergeron, C.R., Prussing, C., Boerlin, P., Daignault, D., Dutil, L., Reid-Smith, R.J., Zhanel, G.G. and Manges, A.R. (2012) Chicken as reservoir for extraintestinal pathogenic *Escherichia coli* in Humans, Canada. *Emerg. Infect. Dis.*, 18(3): 415–421.
- 153. Manges, A.R. (2016) *Escherichia coli* and urinary tract infections: The role of poultry-meat. *Clin. Microbiol. Infect.*, 22(2): 122–129.
- 154. Johnson, T.J., Kariyawasam, S., Wannemuehler, Y., Mangiamele, P., Johnson, S.J., Doetkott, C., Skyberg, J.A., Lynne, A.M., Johnson, J.R. and Nolan, L.K. (2007) The genome sequence of avian pathogenic *Escherichia coli* strain O1:K1:H7 shares strong similarities with human extraintestinal pathogenic *E. coli* genomes. *J. Bacteriol.*, 189(8): 3228–3236.
- 155. Johnson, T.J., Wannemuehler, Y., Johnson, S.J., Stell, A.L., Doetkott, C., Johnson, J.R., Kim, K.S., Spanjaard, L. and Nolan, L.K. (2008) Comparison of extraintestinal pathogenic *Escherichia coli* strains from human and avian sources reveals a mixed subset representing potential zoonotic pathogens. *Appl. Environ. Microbiol.*, 74(22): 7043–7050.
- Manges, A.R. and Johnson, J.R. (2012) Food-borne origins of *Escherichia coli* causing extraintestinal infections. *Clin. Infect. Dis.*, 55(5): 712–719.
- 157. Mellata, M., Johnson, J.R. and Curtiss, R. (2017) *Escherichia coli* isolates from commercial chicken meat and eggs cause sepsis, meningitis and urinary tract infection in rodent models of human infections. *Zoonoses Public Health*, 65(1): 103–113.
- 158. Bauchart, P., Germon, P., Brée, A., Oswald, E., Hacker, J. and Dobrindt, U. (2010) Pathogenomic comparison of human extraintestinal and avian pathogenic *Escherichia coli* search for factors involved in host specificity or zoonotic potential. *Microb. Pathog.*, 49(3): 105–115.
- 159. Lyhs, U., Ikonen, I., Pohjanvirta, T., Raninen, K., Perko-Mäkelä, P. and Pelkonen, S. (2012) Extraintestinal pathogenic *Escherichia coli* in poultry meat products on the Finnish retail market. *Acta Vet. Scand.*, 54(1): 64.
- 160. Ozaki, H. and Murase, T. (2009) Multiple routes of entry for *Escherichia coli* causing colibacillosis in commercial layer chickens. *J. Vet. Med. Sci.*, 71(12): 1685–1689.
- Luhung, Y.G.A., Suarjana, I.G.K. and Gelgel, K.T.P. (2017) Sensitivity isolate *Escherichia coli* pathogen of broiler organs that infected colisepticaemia againts oxytetracycline, ampicillin, and sulfamethoxazole. *Bul. Vet. Udayana*, 9(1): 60–66.
- 162. Lim, J.S., Choi, D.S., Kim, Y.J., Chon, J.W., Kim, H.S., Park, H.J., Moon, J.S., Wee, S.H. and Seo, K.H. (2015) Characterization of *Escherichia coli* producing extended-spectrum β-lactamase (ESBL) isolated from chicken

slaughterhouses in South Korea. Foodborne Pathog. Dis., 12(9): 741–748.

- 163. Wibisono, F.J., Sumiarto, B., Untari, T., Effendi, M.H., Permatasari, D.A. and Witaningrum, A.M. (2020) Short communication: Pattern of antibiotic resistance on extended-spectrum beta-lactamases genes producing *Escherichia coli* on laying hens in Blitar, Indonesia. *Biodiversitas J. Biol. Divers.*, 21(10): 4631–4635.
- Widodo, A., Effendi, M.H. and Khairullah, A.R. (2020) Extendedspectrum beta -lactamase (ESBL)-producing *Eschericia coli* from livestock. *Syst. Rev. Pharm.*, 11(7): 382–392.
- 165. Wibisono, F.J., Sumiarto, B., Untari, T., Effendi, M.H., Permatasari, D.A. and Witaningrum, A.M. (2021) Molecular identification of CTX gene of extended spectrum beta-lactamases (Esbl) producing *Escherichia coli* on layer chicken in Blitar, Indonesia. *J. Anim. Plant Sci.*, 31(4): 6–9.
- 166. Ansharieta, R., Ramandinianto, S.C., Effendi, M.H. and Plumeriastuti, H. (2020) Molecular identification of blactx-m and blatem genes encoding extended-spectrum β-lactamase (ESBL) producing *Escherichia coli* isolated from raw cow's milk in East Java, Indonesia. *Biodiversitas*, 22(4): 1600–1605.
- 167. Rahmahani, J., Salamah, S., Mufasirin, M., Tyasningsih, W. and Effendi, M.H. (2020) Antimicrobial resistance profile of *Escherichia coli* from cloacal swab of domestic chicken in Surabaya traditional market. *Biochem. Cell. Arch.*, 20(1): 2993–2997.
- 168. Harijani, N., Oetama, S.J.T., Soepranianondo, K., Effendi, M.H. and Tyasningsih, W. (2020) Biological

hazard on multidrug resistance (MDR) of *Escherichia coli* collected from cloacal swab of broiler chicken on wet markets Surabaya. *Indian J. Forensic Med. Toxicol.*, 14(4): 3239–3244.

- 169. Wibisono, F.J., Sumiarto, B., Untari, T., Effendi, M.H., Permatasari, D.A. and Witaningrum, A.M. (2020) Antibiotic resistance profile of *Escherichia coli* isolates collected from cloaca swabs on laying hens in Udanawu sub-district, Blitar district, Indonesia. *Ecol. Environ. Conserv.*, 26(Suppl): S261–S264.
- 170. Jeong, J., Lee, J.Y., Kang, M.S., Lee, H.J., Kang, S.I., Lee, O.M., Kwon, Y.K. and Kim, J.H. (2021) Comparative characteristics and zoonotic potential of avian pathogenic *Escherichia coli* (APEC) isolates from chicken and duck in South Korea. *Microorganisms*, 9(5): 946.
- 171. Wibisono, F.M., Faridah, H.D., Wibisono, F.J., Tyasningsih, W., Effendi, M.H., Witaningrum, A.M. and Ugbo, E.N. (2021) Detection of invA virulence gene of multidrug-resistant *Salmonella* species isolated from the cloacal swab of broiler chickens in Blitar district, East Java, Indonesia. *Vet. World*, 14(12): 3126–3131.
- 172. Witaningrum, A.M., Wibisono, F.J., Permatasari, D.A. and Effendi, M.H. (2021) Detection of class 1 integron encoding gene in multidrug resistance (MDR) *Citrobacter freundii* isolated from healthy broiler chicken. *Trop. Anim. Sci. J.*, 44(3): 363–368.
- 173. García, A. and Fox, J.G. (2021) A one health perspective for defining and deciphering *Escherichia coli* pathogenic potential in multiple hosts. *Comp. Med.*, 71(1): 3–45.

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