

Prevalence and antibiotic susceptibility of *Salmonella* spp. from water sources in Tamale, Ghana

Frederick Adzitey, Charles Kwaku Nunana Ashiagbor and Haruna Abu

Department of Animal Science, Faculty of Agriculture, University for Development Studies, Tamale, Ghana.

Corresponding author: Frederick Adzitey, e-mail: adzitey@yahoo.co.uk,

CKNA: cashiabor@yahoo.com, HA: harunaabu81@yahoo.com

Received: 30-06-2016, **Accepted:** 07-09-2016, **Published online:** 16-09-2016

How to cite this article: Adzitey F, Ashiagbor CKN, Abu H. Prevalence and antibiotic susceptibility of *Salmonella* spp. from water sources in Tamale, Ghana. *Int J One Health* 2016;2:24-28.

Abstract

Aim: This study investigated the prevalence and antibiotic resistance of *Salmonella* species isolated from drinking water sources in Tamale Metropolis.

Materials and Methods: Isolation of *Salmonella* species from 275 different drinking water samples (25 each from dam, well, rain, and bottle, 35 from tap, 40 from water trough, and 100 from sachet) was done using a slightly modified method of the Bacteriological Analytical Manual of the Food and Drugs Administration, USA. 34 *Salmonella* species isolated from the water samples were examined for their susceptibility to nine different antibiotics using the disc diffusion method. The study was carried out from July 2014 to January 2015.

Results: The overall prevalence of *Salmonella* species was 4.36% (12/275). Dam 16.00% (4/25) and well 16.00% (4/25) water samples were the most contaminated source, followed by rain water (stored) 12.00% (3/25) and tap water samples 2.86% (1/35). There were no significant differences among water samples which were positive for *Salmonella* species ($p > 0.05$); however, dam and well samples that were positive for *Salmonella* species differ significantly ($p < 0.05$) from bottle water, sachet water, and water trough samples, which were negative for *Salmonella* species. The 34 *Salmonella* isolates were highly resistant to erythromycin (E) (100%) and vancomycin (VA) (94.12%). Few isolates exhibited intermediate resistances to ceftriaxone (CRO) (17.65%), gentamicin (CN) (17.65%), tetracycline (14.71%), chloramphenicol (C) (5.88%), ciprofloxacin (CIP) (2.94%), and amoxicillin (AMC) (2.94%). *Salmonella* isolates also exhibited six different antibiotic resistant patterns (VA-E, VA-E-AMC, VA-E-CRO, VA-E-C, VA-E-CRO-AMC, and VA-E-AMC-CN). The resistant pattern VA-E (with multiple antibiotic resistance index of 0.22) was the commonest.

Conclusion: This study indicated that some drinking water sources for humans and animals in Tamale Metropolis are contaminated with *Salmonella* species which exhibited varying resistance to various antibiotics. Therefore, consumers of water at the Tamale Metropolis are at risk of *Salmonella* infection from drinking water from positive water sources in the Tamale Metropolis.

Keywords: antibiotics, drinking water, public health, *Salmonella* species.

Introduction

Water is very essential for the existence of humans and other forms of life on earth [1]. It is involved in the normal functioning of cells that make up humans and all living organisms [2,3]. Water also helps to remove waste, lubricates joints, serve as a major component of blood, and a necessary medium for many chemical reactions that help in the formation of meat, eggs, and other animal and plant foods [4]. Potable water is expected to be odorless, tasteless, free from toxic substances, microbial and chemical contaminations [2,5]. Water is also known to be the dwelling place for most bacteria and other microorganisms which cause a variety of waterborne infections such as cholera, typhoid, bilharzia, and malaria [6,7].

Salmonella species are Gram-negative facultative anaerobe bacteria and have been isolated from humans, animals, and the environment [8-14]. *Salmonellae* are among the major pathogenic bacteria in humans as well as in animals [15,16]. They are the leading cause of acute gastroenteritis in several countries and salmonellosis remains an important public health problem worldwide [15]. In Canada, 8.5% (342) of river water samples were contaminated with *Salmonella* species which were influenced by rainfall runoff and drainage from agricultural land [17]. In Czech, 16% (87) of pond water samples were contaminated with *Salmonella* species [18]. In South Africa, the prevalence rate of *Salmonella* species in different river samples ranged from 33-90% [19]. A study by Akinyemi *et al.* [20] in Nigeria indicated that *Salmonella* was present in water samples. Hitherto published work on the isolation and antibiotic resistance of *Salmonella* species in Ghana appears to be unavailable. Antibiotics continue to play a very important role in decreasing diseases, illness and/or death associated with bacterial infections [21]. Human activities have been largely linked to the emergence of multidrug resistance isolates [16,22].

Copyright: Adzitey, *et al.* This article is an open access article distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated.

Salmonella species isolated from water samples were resistant to 2 or more antibiotics [21]. This study was conducted to find out whether drinking water sources in the Tamale, Metropolis of Ghana are contaminated with *Salmonella* species. The study also investigated the antibiotic resistance of the *Salmonella* species isolated from the water samples.

Materials and Methods

Ethical approval

All samples were collected as per the standard sample collection procedure without harming humans or animals.

Location of study

The study was conducted in Tamale Metropolis. Tamale is the capital town of the Northern Region of Ghana. It is located within the Guinea Savannah belt. Tamale is the third most populous settlement in Ghana, in terms of population, with a population of 537,986 people [23]. Tamale Metropolis can be located on longitude 09°24'27" North and latitude 00°51'12" West [23]. The Tamale Metropolis occupies approximately 750 km², which is 13% of the total area of the Northern Region [23].

Sample collection

A total of 275 water samples were sampled. These water samples were collected aseptically from dam (n=25), well (n=25), rain (n=25), tap (n = 35), bottle (n=25), sachet (n=100), and water trough (n=40). The water samples were stored at 4°C and transported to the Spanish Laboratory of University for Development Studies, Nyankpala, Ghana, where analysis was carried out immediately upon arrival for the presence of *Salmonella* species.

Bacteriological analysis

Analyses for *Salmonella* species were done using a modified method according to the Bacteriological Analytical Manual of the Food and Drug Administration [8]. 500 ml of the various water samples were obtained and thoroughly mixed. 1 ml of water was taken from the 500 ml, transferred into 10 ml buffered peptone water, and incubated at 37°C for 24±2 h. Afterward, 0.1 ml and 1 ml of pre-enriched aliquots were transferred into 10 ml Rappaport and Vassiliadis broth and 10 ml selenite cystine broth, respectively. Enrichment samples in Rappaport and Vassiliadis broth were incubated at 42°C for 24 h while that of the selenite cystine broth were incubated at 37°C for 24 h. Enriched aliquots (ca. 10 µl) were then streaked onto xylose lysine deoxycholate and brilliant green bile agar and incubated at 37°C for 24-48 h. Presumptive *Salmonella* species were purified on nutrient agar and were identified and confirmed using Gram-staining, biochemical tests, and *Salmonella* latex agglutination test. All incubations were done under aerobic condition, and all media used were purchased from Oxoid, Oxoid Limited, Basingstoke, UK.

Antimicrobial susceptibility of *Salmonella* species

The disk diffusion method of Bauer *et al.* [24] was used to determine the antibiotic resistance of 34 *Salmonella* species against the following antibiotics: Amoxicillin/clavulanic acid (AMC) 30 µg, chloramphenicol (C) 30 µg, gentamicin (CN) 10 µg, ceftriaxone (CRO) 30 µg, ciprofloxacin (CIP) 5 µg, erythromycin (E) 15 µg, sulfamethoxazole/trimethoprim (SXT) 22 µg, tetracycline (TE) 30 µg, and vancomycin (VA) 30 µg. The disks were purchased from Oxoid Limited, Basingstoke, UK. Pure cultures of *Salmonella* species were grown overnight in tryptic soy broth (TSB) (Oxoid Limited, Basingstoke, UK) at 37°C and the concentration adjusted using sterile TSB until a 0.5 McFarland turbidity was attained. About 100 µl of the culture was then swabbed onto Mueller-Hinton agar (Oxoid Limited, Basingstoke, UK) using a sterile cotton swab. Three antimicrobial disks were placed on the surface of the agar plate at a distance to avoid overlapping of inhibition zones. The plates were incubated at 37°C for 16-18 h, and the results were interpreted as sensitive, intermediate, or resistance according to the Clinical and Laboratory Standards Institute Guidelines [25]. The multiple antibiotic resistance (MAR) index was calculated and interpreted according to Krumperman [26] using the formula: a/b, where "a" represents the number of antibiotics to which a particular isolate was resistant and "b" the total number of antibiotics tested.

Statistical analysis

The data obtained were analyzed using the generalized linear model of Statistical Package for the Social Sciences (SPSS) Version 17.

Results

The results for the occurrence of *Salmonella* species in water samples examined in the Tamale Metropolis are presented in Table-1. The overall prevalence of *Salmonella* species was 6.36% (12/275). Dam (16.00%) and well (16.00%) water samples were the most contaminated source, followed by rain water (12.00%) and tap water samples (2.86%). Bottle, sachet, and water trough samples recorded 0.00% (0/25), (0/100), and (0/40), respectively. There were no significant differences among water samples which were positive for *Salmonella* species (p>0.05);

Table-1: Distribution of *Salmonella* species in the various drinking water sources tested.

| Sources of drinking water | Number of samples tested | Number of samples positive | Percentage prevalence |
|---------------------------|--------------------------|----------------------------|-----------------------|
| Dam water | 25 | 4 | 16.00 |
| Well water | 25 | 4 | 16.00 |
| Rain water | 25 | 3 | 12.00 |
| Tap water | 35 | 1 | 2.85 |
| Bottle water | 25 | 0 | 0.00 |
| Sachet water | 100 | 0 | 0.00 |
| Water trough | 40 | 0 | 0.00 |
| Total | 275 | 12 | 4.36 |

however, dam and well water samples that were positive for *Salmonella* species differ significantly ($p < 0.05$) from bottle water, sachet water, and water trough samples.

The antibiotic susceptibility test of the *Salmonella* isolates is shown in Table-2. Resistant to E and VA was 100% and 94.12%, respectively. None of the *Salmonella* isolates was resistant to SXT, TE, and CIP, even though intermediate resistances occurred for TE (14.71%) and CIP (2.94%). Most of the isolates were highly susceptible to C (91.18%), CN (79.41%), AMC acid (73.53%), and CRO (70.59%).

The 34 *Salmonella* isolates also exhibited six different antibiotic resistant patterns. 23 of the isolates exhibited a resistant pattern of VA-E with MAR index of 0.22, five exhibited a pattern of VA-E-AMC with MAR index of 0.33, two isolates exhibited a pattern of VA-E-CRO with MAR index of 0.33, one exhibited a pattern of VA-E-C with a MAR index of 0.33, two exhibited a pattern of VA-E-CRO-AMC with MAR index of 0.44, and one exhibited a pattern of VA-E-AMC-CN with MAR index of 0.44 (Table-3).

Discussion

Of the 275 different drinking water samples examined, 12 were positive for *Salmonella* species. The occurrence of *Salmonella* species was highest in dam and well water samples, followed by rain and tap water samples. The presence of *Salmonella* species in dam, well, rain, and tap water samples is worrying since the majority of the people and animals in the Tamale Metropolis, and its environs drink from them. It was observed that dams which were positive for *Salmonella* species were closer to human settlement and under high utilization by humans and animals. Some of the wells which were contaminated with *Salmonella* species were shallow, without proper cover and the containers used for fetching water from them were dirty and left on the ground. In this study, we collected rain water from stored containers/tanks and directly from the sky and roof (aluminum sheet) during raining. Water samples collected directly from the roof and the sky were negative for *Salmonella* species. Rainwater samples that were contaminated with *Salmonella* species were collected from storage containers/tanks. Containers used for fetching water from these storage containers/tanks were also

sometimes dirty, left on the ground and reuse without washing. The containers used for fetching water from wells and storage tanks can easily get contaminated by soil and/or feces of domestic animals which were observed roaming around the wells and storage tanks in search of water. Ghana Water Company supplies water (tap water) to the various households in Tamale and its environs. This company ensures that the water supplied to consumers is of good quality without any contamination. One tap water sample collected from a chop bar was contaminated with *Salmonella* species. This contamination may be as a result of unhygienic handling of tap and personal hygiene of consumers. Indiscriminate defecation by humans and animals in and around water bodies can cause contamination of drinking water sources such as dams and wells. Dirts, chemicals, and/or feces created by humans and/or animal activities around water bodies can be washed back into these water bodies (dams and wells), especially by rains. Variation in the level of microorganisms in water bodies can be attributed to indiscriminate human and animal feces around such water bodies [27]. *Salmonella* species were not isolated from bottle water (four different brands), sachet water (four different brands), and water trough (from ruminant and poultry farms). This indicates that the bottle and sachet water samples were treated and packaged under good sanitary conditions without fecal or environmental contamination. The World Health Organization [28] reported that good quality drinking water should be free from microbial contamination. The absence of *Salmonella* species in the water trough samples could be attributed to the fact that appropriate management practices are being carried out at the farms visited. Frequent change of water and cleaning of water troughs create a good hygienic and sanitary condition which will prevent bacteria contamination and growth.

Around 34 *Salmonella* species isolated from the drinking water sources in the Tamale Metropolis were examined for their susceptibility to nine different antibiotics. They were highly resistant to E (100.00%) and VA (94.12%). Resistance to AMC was 23.53%. Few isolates also exhibited intermediate resistances to CRO (17.65%), CN (17.65%), TE (14.71%), C (5.88%), CIP (2.94%), and AMC (2.94%). Intermediate resistance means those *Salmonella* species were not

Table-2: Antibiotic susceptibility of drinking water *Salmonella* isolates.

| Antibiotics | % Resistant | % Intermediate | % Susceptibility |
|-------------------------------|-------------|----------------|------------------|
| Chloramphenicol | 2.94 | 5.88 | 91.18 |
| Gentamicin | 2.94 | 17.65 | 79.41 |
| Vancomycin | 94.12 | 0.00 | 5.88 |
| Sulfamethoxazole/trimethoprim | 0.00 | 0.00 | 100.00 |
| Tetracycline | 0.00 | 14.71 | 85.29 |
| Erythromycin | 100.00 | 0.00 | 0.00 |
| Ciprofloxacin | 0.00 | 2.94 | 97.06 |
| Ceftriaxone | 11.76 | 17.65 | 70.59 |
| Amoxicillin/clavulanic acid | 23.53 | 2.94 | 73.53 |

Table-3: Antibiotic resistance pattern of the *Salmonella* species.

| Code | Sources | Antibiotic resistance profile | MAR index |
|------|--------------------|-------------------------------|-----------|
| D1 | Dam water | VA-E | 0.22 |
| D2 | Dam water | VA-E | 0.22 |
| D3 | Dam water | VA-E | 0.22 |
| D4 | Dam water | VA-E | 0.22 |
| D5 | Dam water | VA-E | 0.22 |
| D6 | Dam water | VA-E | 0.22 |
| S1 | Storage rain water | VA-E | 0.22 |
| S2 | Storage rain water | VA-E | 0.22 |
| T1 | Tap | VA-E | 0.22 |
| T2 | Tap | VA-E | 0.22 |
| T3 | Tap | VA-E | 0.22 |
| T4 | Tap | VA-E | 0.22 |
| T5 | Tap | VA-E | 0.22 |
| T6 | Tap | VA-E | 0.22 |
| D7 | Dam water | VA-E | 0.22 |
| D8 | Dam water | VA-E | 0.22 |
| D9 | Dam water | VA-E | 0.22 |
| D10 | Dam water | VA-E | 0.22 |
| D11 | Dam water | VA-E | 0.22 |
| S3 | Storage rain water | VA-E | 0.22 |
| S4 | Storage rain water | VA-E | 0.22 |
| S5 | Storage rain water | VA-E | 0.22 |
| S6 | Storage rain water | VA-E | 0.22 |
| W1 | Well water | VA-E-AMC | 0.33 |
| W2 | Well water | VA-E-AMC | 0.33 |
| W3 | Well water | VA-E-AMC | 0.33 |
| W4 | Well water | VA-E-AMC | 0.33 |
| S7 | Storage rain water | VA-E-AMC | 0.33 |
| D12 | Dam water | VA-E-CRO | 0.33 |
| D13 | Dam water | VA-E-CRO | 0.33 |
| D14 | Dam water | VA-E-C | 0.33 |
| W5 | Well water | VA-E-CRO-AMC | 0.44 |
| S8 | Storage rain water | VA-E-CRO-AMC | 0.44 |
| W6 | Well water | VA-E-AMC-CN | 0.44 |

C=Chloramphenicol, CN=Gentamicin, VA=Vancomycin, E=Erythromycin, CRO=Ceftriaxone, AMC=Amoxicillin/clavulanic acid, MAR=Multiple antibiotic resistance

clearly resistant or susceptible, and such isolates have the tendency to easily become resistant [29,30]. It has also been indicated that in clinical diagnoses patients with intermediate results can be given a higher dosage of antibiotics [29,30]. The 34 *Salmonella* isolates also exhibited six different antibiotic resistant patterns. The patterns were VA-E, VA-E-AMC, VA-E-CRO, VA-E-C, VA-E-CRO-AMC, and VA-E-AMC-CN. The resistant pattern VA-E was the most common and was exhibited by 23 *Salmonella* isolates. Resistant to as high as four different antibiotics (VA-E-CRO-AMC and VA-E-AMC-CN) was exhibited by 3 *Salmonella* isolates. *Salmonella* species are important pathogens of global concern, and the emergence of antibiotic resistant strains is largely due to indiscriminate use of antibiotics in animal feeds as growth promoters and as therapeutic agents for treating humans and animals [31].

Conclusion

This study indicated that some drinking water sources for humans and animals in Tamale Metropolis

are contaminated with *Salmonella* species. In addition, they are resistant to some antibiotics at different rates. Therefore, drinking water sources in Tamale Metropolis are potential sources for the transmission of *Salmonella* infection.

Authors' Contributions

FA designed the experiment. CKNA and HA performed all the experiments. FA analyzed the data and wrote the draft manuscript. All authors read and approved the final manuscript.

Acknowledgments

The authors are gratefully to the University for Development Studies, for allowing them to use the Spanish Laboratory to carry out this experiment. No fund was received from any institute for this study.

Competing Interests

The authors declare that they have no competing interests.

References

- Adzitey F, Sumaila N, Saba CK. Isolation of *E. coli* from drinking water sources for humans and farm animals in Nyankpala Community of Ghana. *Res J Microbiol* 2015a;10:126-31.
- Maheshwari N. *Clinical Microbiology and Parasitology*. 2nd ed. New Delhi, India: Jaypee Brothers Medical Publishers; 2008. p. 272.
- Sobsey MD, Bartram S. Water quality and health in the new millennium: The role of the World Health Organization Guidelines for Drinking-Water Quality. *Forum Nutr* 2003;56:396-405.
- Organic Nutrition. The benefits of drinking water. Available from <https://www.organicnutrition.co.uk/articles/benefits-of-drinking-water.htm>. Accessed on 01-05-2016.
- Adzitey F, Nafisah S, Haruna A. Antibiotic susceptibility of *E. coli* isolated from some drinking water sources in Tamale Metropolis of Ghana. *Curr Res Bacteriol* 2015b; 8:34-40.
- Spellman FR, Drinan J. *The Drinking Water Handbook*. Lancaster, Pennsylvania, USA: Technomic Publishing Co., Inc.; 2000. p. 260.
- World Health Organization. In: Cotruvo JA, Dufour A, Rees G, Bartram J, Carr R, Cliver DO, *et al.*, editors. *Waterborne Zoonosis: Identification, Causes and Control*. London, UK: IWA Publishing; 2004. p. 95-111.
- Wallace HA, Hammack TS. *Salmonella* in *Bacteriological Analytical Manual*. Available from: <http://www.fda.gov/Food/ScienceResearch/LaboratoryMethods/BacteriologicalAnalyticalManualBAM/UCM070149>. Accessed on 01-06-2014.
- Frederick A, Huda N. *Salmonellas*, poultry house environments and feeds: A review. *J Anim Vet Adv* 2011;10:679-85.
- Adzitey F, Nsoah JK, Teye G. Prevalence and antibiotic susceptibility of *Salmonella* species isolated from beef and its related samples in Techiman Municipality of Ghana. *Turk J Agric Food Sci Technol* 2015c;3:644-50.
- Anachinaba IA, Adzitey F, Teye GA. Assessment of the microbial quality of locally produced meat (beef and pork) in Bolgatanga Municipal of Ghana. *Internet J Food Saf* 2015;17:1-5.
- Makwana PP, Nayak JB, Brahmabhatt MN, Chaudhary JH. Detection of *Salmonella* spp. from chevon, mutton and its environment in retail meat shops in Anand city (Gujarat), India. *Vet World* 2015;8:388-92.
- Kalambhe DG, Zade NN, Chaudhari SP, Shinde SV,

- Khan W, Patil AR. Isolation, antibiogram and pathogenicity of *Salmonella* spp. recovered from slaughtered food animals in Nagpur region of Central India. *Vet World* 2016;9:176-81.
14. Papadopoulos T, Zdragas A, Mandilara G, Vafeas G, Giantzi V, Petridou E, *et al.* Characterization of *Salmonella* isolates from municipal sewage, patients, foods, and animals in Greece using antimicrobial susceptibility testing and pulsed field gel electrophoresis. *Int J One Health* 2016;2:12-8.
 15. Addis Z, Kebede N, Worku Z, Gezahegn H, Yirsaw A, Kassa T. Prevalence and antimicrobial resistance of *Salmonella* isolated from lactating cows and in contact humans in dairy farms of Addis Ababa: A cross sectional study. *BMC Infect Dis* 2011;11:222.
 16. Wegener HC. Antibiotic resistance linking human and animal health. In: *Improving Food Safety Through a One Health Approach: Workshop Summary*. Washington, DC: National Academies Press; 2012. p. 331-49. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK114485/>. Accessed on 18-02-2014.
 17. Jokinen C, Edge TA, Ho S, Koning W, Laing C, Mauro W, *et al.* Molecular subtypes of *Campylobacter* spp. *Salmonella enterica*, and *Escherichia coli* O157:H7 isolated from faecal and surface water samples in the Oldman River Watershed, Alberta, Canada. *Water Res* 2011;45:1247-57.
 18. Dolejská M, Bierosová B, Kohoutová L, Literák I, Cízek A. Antibiotic-resistant *Salmonella* and *Escherichia coli* isolates with integrons and extended-spectrum beta-lactamases in surface water and sympatric black-headed gulls. *J Appl Microbiol* 2009;106:1941-50.
 19. Obi CL, Potgieter N, Musie EM, Igumbor EO, Bessong PO, Samie A, *et al.* Human and environmental-associated non-typhoidal *Salmonella* isolates from different sources in the Venda Region of South Africa. *Proceedings of the 2004 Water Institute of Southern Africa (WISA) Biennial Conference*. 2004. p. 51-7.
 20. Akinyemi KO, Iwalokun BA, Foli F, Oshodi K, Coker AO. Prevalence of multiple drug resistance and screening of enterotoxin (stn) gene in *Salmonella enterica* serovars from water sources in Lagos, Nigeria. *Public Health* 2011;125:65-71.
 21. Adzitey F. Antibiotic classes and antibiotic susceptibility of bacterial isolates from selected poultry; A mini review. *World's Vet J* 2015a;5:36-41.
 22. Abakpa GO, Umoh VJ, Ameh JB, Yakubu SE, Kwaga JK, Kamaruzaman S. Diversity and antimicrobial resistance of *Salmonella enterica* isolated from fresh produce and environmental samples. *Environ Nanotech Monitory Mgt* 2015;3:38-46.
 23. Anonymous. Tamale Metropolis. Available from: http://www.en.wikipedia.org/wiki/Tamale_Ghana; 2014. Accessed on 18-02-2014.
 24. Bauer AW, Kirby WM, Sherris JC, Turck M. Antibiotic susceptibility testing by a standardized single disk method. *Am J Clin Pathol* 1966;45:493-6.
 25. Clinical and Laboratory Standards Institute. *Performance Standards for Antimicrobial Disk and Dilution Susceptibility Tests for Bacteria Isolated from Animals Approved Standard CLSI Document VET01-A4*. In *Approved Standard*. 4th ed. Wayne, PA, USA: CLSI; 2013.
 26. Krumpferman PH. Multiple antibiotic resistance indexing of *Escherichia coli* to identify high-risk sources of fecal contamination of foods. *Appl Environ Microbiol* 1983;46:165-70.
 27. Adentunde LA, Glover RL. Evaluation of bacteriological quality of drinking water used by selected secondary schools in Navorongo in Kassina-Nankana District in the Upper East Region of Ghana. *Prim J Microbiol Res* 2011;1:47-51.
 28. World Health Organization. *Microbial Aspect in Guideline for Drinking Water Quality*. 3rd ed. Geneva: WHO; 2006.
 29. Adzitey F. Antibiotic resistance of *Escherichia coli* isolated from beef and its related samples in Techiman Municipality of Ghana. *Asian J Anim Sci* 2015b;9:233-40.
 30. Adzitey F, Rusul G, Huda N, Cogan T, Corry J. Prevalence, antibiotic resistance and RAPD typing of *Campylobacter* species isolated from ducks, their rearing and processing environments in Penang, Malaysia. *Int J Food Microbiol* 2012;154:197-205.
 31. Plym FL, Wierup M. *Salmonella* contamination: A significant challenge to the global marketing of animal food products. *Rev Sci Tech* 2006;25:541-54.
